

A Multi-Frame Optical Flow Spot Tracker

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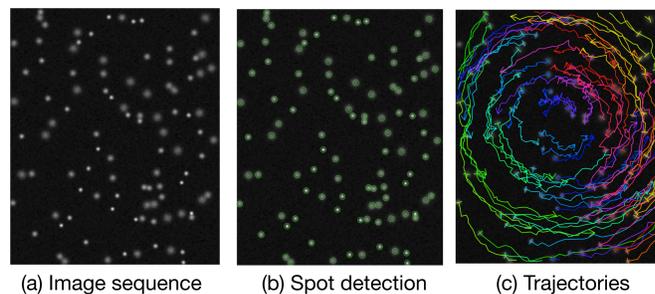


Summary

Accurate and robust spot tracking is a necessary tool for quantitative motion analysis in fluorescence microscopy images. In this work, we exploit **the underlying stationary motion** in biological systems, e.g. the movement of crowds, bacteria swarming and cyclosis in plant cells, and then propose a multi-frame optical flow based tracker. We obtain the stationary motion by adapting a recent optical flow algorithm that relates one image to another locally using an all-pass filter. We perform this operation over all the image frames simultaneously and estimate a single, stationary optical flow. We compare the proposed tracker with two existing techniques and show that our approach is more robust to high noise and varying structure. In addition, we also show initial experiments on real microscopy images.

Spot Tracking

Problem: To detect and track a set of spots over an image sequence comprising N frames.



Our Approach

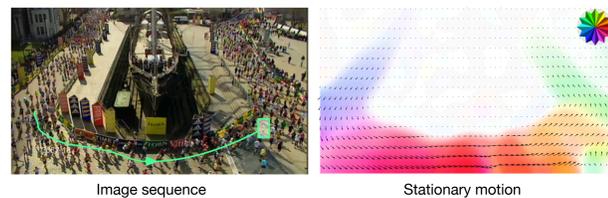
(a) Stationary Motion

Assumption: there exists a motion pattern that underpins the dynamics within the system being imaged, called *the stationary motion field*.

This underlying motion is the main part of the motion between two consecutive images that is consistent across all the images in the sequence.

This type of motion has been observed in many organisms:

- Crowds of people
- Flocks of birds
- Bacteria
- Plant cells

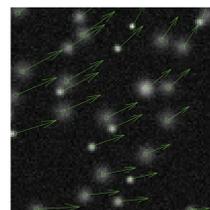


(b) Stationary Local All-Pass (SLAP) Tracker

Step 1: Estimation of an underlying stationary motion field over all image frames simultaneously using an adapted version of the local all-pass algorithm [1].

Step 2: Simple spot detection.

Step 3: Trajectory estimation based on the estimated stationary motion filed.



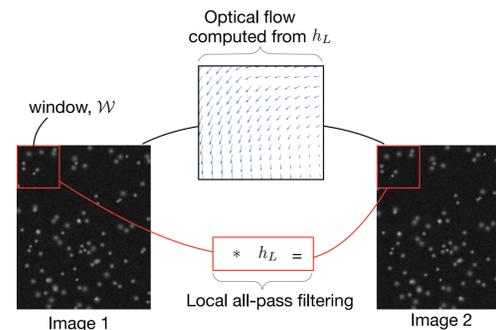
↔ Use the stationary motion to guide the spot tracking

Stationary Motion Estimation

1. Motion estimation using Local All-Pass (LAP) filters

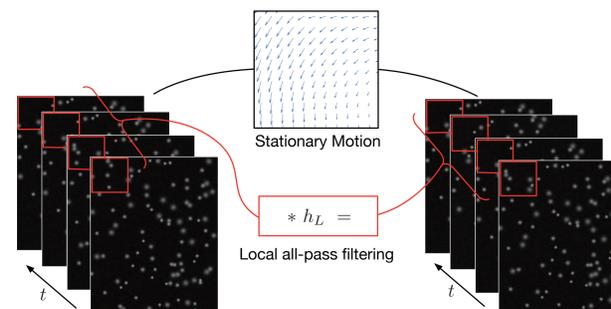
The LAP algorithm proposed in [1]:

- Constant motion \implies All-pass filtering
- Relate local windows in two images using this filter
- Extract the motion vectors from the local all-pass filters



2. Adaption to stationary motion

We estimate one single motion field from the whole image sequence simultaneously.

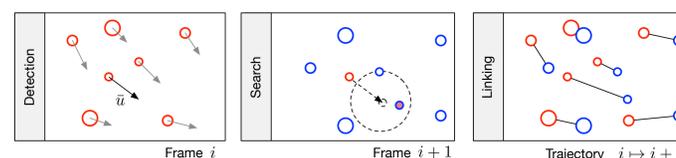


↔ Extract the stationary motion field \bar{u} from these filters.

Detection and Trajectory Estimation

Spots Detection: The spot locations are estimated by finding the local maxima of the correlations with a pre-defined Gaussian mask.

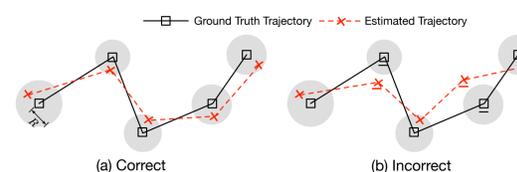
Trajectory Estimation: The trajectory is built based on associating the spot in the next frame that is closest to the predicted position.



Experiment and Results

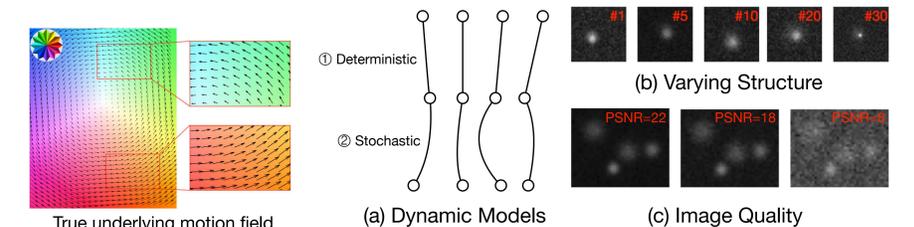
Performance Evaluation:

We evaluate a tracker based upon the percentage of correctly identified trajectories.



Experiment and Results

Synthetic Images: The synthetic image sequence consists of two elements: the spots we wish to track and the underlying stationary motion that controls their movement.

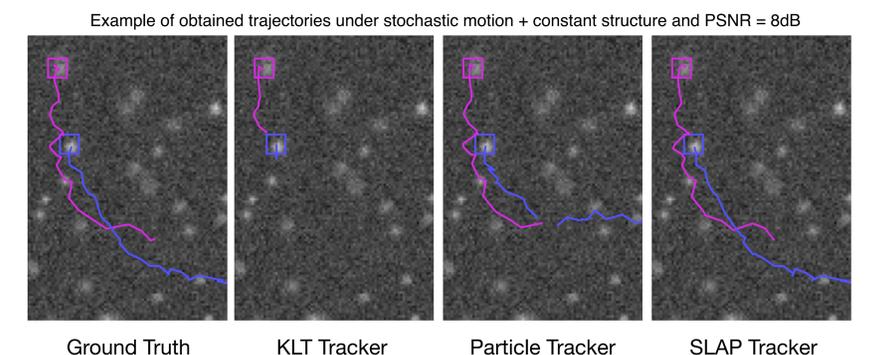


Results:

Scenarios	KLT Tracker [2]			Particle Tracker [3]			SLAP Tracker		
	PSNR=22	PSNR=18	PSNR=8	PSNR=22	PSNR=18	PSNR=8	PSNR=22	PSNR=18	PSNR=8
Deterministic+Constant	56%	56%	53%	75%	74%	57%	90%	87%	76%
Deterministic+Varying	51%	54%	47%	67%	63%	48%	80%	75%	62%
Stochastic+Constant	29%	28%	23%	39%	38%	31%	62%	60%	50%
Stochastic+Varying	27%	24%	18%	38%	29%	23%	59%	57%	45%

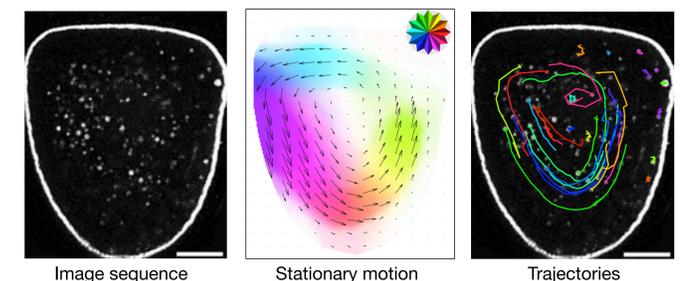
↔ The simple detection and linking strategies have low computational cost.

↔ The stationary motion field is only calculated once across all the images using a very fast algorithm [1], thus it is independent of the spot number.



Real Images:

- The sequence depicts cytoplasmic streaming in *Drosophila oocytes* (fruit fly eggs) [4].
- The SLAP Tracker successfully followed 25 spots (correctness = 83.3%) during 30 consequent frames until their disappearance.



References

- [1] C. Gilliam and T. Blu, "Local all-pass filters for optical flow estimation," in *Proc. IEEE Int. Conf. Acoust. Speech Signal Process. (ICASSP)*, Brisbane, Australia, pp. 1533-1537, April 19-24, 2015.
- [2] J. Shi and C. Tomasi, "Good features to track," *Proc. Computer Vision and Pattern Recognition (CVPR)*, Seattle, Washington, pp. 593-600, June, 1994.
- [3] I. Sbalzarini and P. Koumoutsakos, "Feature point tracking and trajectory analysis for video imaging in cell biology," *J. Struct. Biol.*, vol. 151, no. 2, pp. 182-195, 2005.
- [4] L. Serbus et al., "Dynein and the actin cytoskeleton control kinesin-driven cytoplasmic streaming in *drosophila oocytes*," *Development*, vol. 132, no. 16, pp. 3743-3752, 2005.

